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# Validation of 4 ELISA kits for the screening of nitrofuran metabolites (AOZ, AMOZ, SEM, AHD) in porcine muscle

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# **SUMMARY**

- The use of nitrofurans (furaltadone, nitrofurantoin and nitrofurazone) has been banned for food producing animals since 1993 and since 1995 for furazolidone. Nitrofuran parent drugs are very quickly metabolised. The main protein-bound metabolites are 3-amino-5morpholinomethyl-2-oxazolidinone (AMOZ), 1-aminohydantoin (AHD), semicarbazide (SEM). and3-amino-2-oxazolidinone (AOZ), respectively.
- Since beginning of 2000's, the analysis of residues of nitrofuran drugs is based on the detection of the protein-bound metabolites. In 2003, a definitive MRPL (Minimum Required Performance Limit) was set at 1.0 µg/kg in the European Union for nitrofuran metabolites in poultry and aquaculture products. Since that date, costly LC/MS-MS methods have been developed for the screening and the confirmation of nitrofuran metabolites in line with EU regulations.



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Until last year, only two ELISA kits were commercialized for the screening of AOZ and AMOZ. However, 4 ELISA kits have been recently developed and commercialized for the detection of the 4 nitrofuran metabolites (AOZ, AMOZ, SEM and AHD). These kits have been evaluated and validated in our laboratory according to European Decision 2002/657/EC [1] which concerns the performance of analytical methods. Different performance characteristics (specificity, detection capabilities and cross-reactions) have been determined for each kit.



## **Material and methods**

Preparation of porcine muscle:	<b>ELISA procedures:</b>	Validation protocol:		
1g ± 0.05g of the	Add 50 µl of extract or 50 µl of standard	The analyses have been performed blindly (random number given to samples).		
homogenized sample	solution into each well,	Charificity, false necitive rates Canacity of a method to dictinguish the measured analyte		
$\checkmark$	√ Add 50 ul of the entitledy colution	- <u>Specificity, faise positive rate.</u> Capacity of a method to distinguish the measured analyte		
4 ml deionized water, 0.5 ml 1 M HCl, 100 µl	seal the microplate	Trom other substances.		
10 mM 2-Nitrobenzaldehyde solution, shake	$\downarrow$	Analyse in duplicate $n > or = 60$ representative blank samples.		
$\downarrow$	Incubate at 37°C for 30 min	- Detection capability CCB: CCB = the concentration level. where only $\leq$ 5 % false		
Incubate at 37°C over night		compliant results remain (maximum 2 false compliant out of 60 fortified camples)		
	Add 250 µl/well of washing bufferfor 15 sec	compliant results remain (maximum 5 laise compliant out of ob fortheu samples).		
5ml 0.1 M K2HPO4, 0.4ml 1M NaOH,	Popoat 5 times	Analyse in duplicate $n > or = 60$ blank samples spiked at the MRPL (1 µg/kg) or lower with		
5ml ethyl acetate, shake 30s		the target molecule of each kit.		
$\checkmark$	Add 100 µl enzyme conjugate	- Cross-reactions: Molecules at 10000 ng/g in duplicate: 4 parent nitrofuran molecules		
Centrifuge at 3500 g at room	Incubate at 37°C for 30 min	(furzzelidene, furzltzdene, pitrofurzzene, pitrofurzntein), the 2 other metzbeliter		
temperature for 10 min	$\checkmark$	(Turazoliuone, Turaitauone, Titrorurazone, Titrorurantoin), the 5 other metabolites		
$\checkmark$	Add 250 µl/well of washing bufferfor 15 sec	depending on the kit + 6 major categories: beta-lactams, sulfonamides, tetracyclines,		
Transfer 2.5ml ethyl acetate layer (upper layer)	Take out and flap to dry with absorbent paper	macrolides, aminoglycosides, quinolones.		
Evaporate to dryness by nitrogen at 50°C	Repeat 5 times	- Precision (intra and inter-day repeatability). Spiked samples at 1 1 5 and 2 times the		
↓	↓ Add 50 ul of the cubetrate A	MDDL Analyse at least 6 replicate (3 days)		
Dissolve dry residue in 1ml N-hexane,	solution and 50 µl of the Risolution	MIREL ANALYSE AL LEAST O TEPHCALE (3 UAYS).		
Add Timi of diluted redissolving solution, mix	I.			

temperature for 10min ↓ Take 50µl of the lower aqueous phase per well in the assay <i>Fold of dilution of the sample:2</i>	Incubate at 37°C for 15 min at dark ↓ Add 50 µl of the stop solution. Mix gently ↓ Read at 450 nm (Recommended dual-wavelength 450/630 nm)	An international proficiency test for the screening and the confirmation of nitrofuran metabolites in porcine muscle was held during March-April 2008 by the Community Reference Laboratory (CRL). We have participated with the 4 ELISA kits for nitrofuran metabolites, to complete the validation study.
	Mix gently Incubiets, et 27°C for 45 raise et derts	Participation to an international proficiency test (IPT)
	<ul> <li>Centrifuge at 3500 g at room temperature for 10min</li> <li>↓</li> <li>Take 50µl of the lower aqueous phase per well in the assay</li> <li>Fold of dilution of the sample:2</li> </ul>	<ul> <li>✓ Centrifuge at 3500 g at room temperature for 10min</li> <li>✓ Take 50µl of the lower aqueous phase per well in the assay</li> <li><i>Fold of dilution of the sample:2</i></li> <li>Mix gently Incubate at 37°C for 15 min at dark</li> <li>✓ Add 50 µl of the stop solution. Mix gently</li> <li>✓ Read at 450 nm (Recommended dual-wavelength 450/630 nm)</li> </ul>

# NESUILS

OD<0,55

# **Specificity / Detection capability: Example of AMOZ kit:**



# **Example of AHD kit:**

**Distribution of the Optical Densities of blank and** samples spiked at MRPL with AHD



1.03 OD

Cut-off AHD

Number of false +

Number of false -

The 4 ELISA kits showed no cross-reactions (< 0.01 %) with the other nitrofuran metabolites and with other families of families. However, the 4 ELISA kit for each metabolite showed variable cross-reactions with the parent drug.

ELISA kit for	Parent Drug	Cross-reactions (%)
AOZ	Furazolidone	1.5
AMOZ	Furaltadone	42
AHD	Nitrofurantoine	0.1
SEM	Nitrofurazone	13

# **Precision**

**0,34 conc** 

11

conc

19

The variability of the response in OD was smaller than the variability of the calculated concentrations for the AHD kit, especially inter-day variability.



**Distribution of the calculated concentrations of blank** and samples spiked at MRPL with AMOZ

if OD<0,77 conc>0,6 ng/g conc>0,75 ng/g



The better cut-off value is the one where the false negative rate is lower than 5 % and the false positive rate as low as possible. For the screening of <u>AMOZ</u>, the best choice was to set the cut-off at 0.75 ng/g for the calculated concentration or at 0.77 OD unit (mean OD spiked + 2.33\*SD). For the screening of <u>AHD</u>, the best choice was to set the cut-off at 0.34 ng/g for the calculated concentration or at 1.1 OD unit (mean OD blank -1.64\*SD).

1,1 OD

11

# **International Proficiency Test**

The results were satisfactory for AHD. The results for AOZ were satisfactory if the cut-off was set in concentration. The results for AMOZ and SEM were unsatisfactory.



# Conclusions

The overall conclusion of this validation study is that the ELISA kit for the screening of AHD is very interesting, regarding its performance and knowing that it is the first commercial kit available for the screening of AHD in food matrices. It should be interesting to test this kit on other matrices (milk, shrimp, fish, honey). The ELISA kit for the screening of SEM is also a new offer. Unfortunately, its performance was unsatisfactory. It was impossible to set a reasonable cut-off value which should be a compromise between the false negative and the false positive rate. Moreover, the participation to the IPT gave many false positive results. The results of the 2 ELISA kits for the screening for AMOZ and AOZ kit were unsatisfactory with high false positive rates.

### References

[1] Decision 2002/657/EC of 12 August 2002, implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results, Off. J. European Union L221 (2002) 8–36. [2] V. Gaudin, C. Hedou, P. Sanders, Validation of a Biacore method for screening eight sulphonamides in milk and porcine muscle tissues according to European decision 2002/657/EC. Journal of AOAC, 90 (6) (2007) 1706-1715.