



**anses**



**Nancy Laboratory for Rabies and  
Wildlife**

**European Union Reference Laboratory**

**UNIT : Lyssavirus**

**for Rabies**

**PROFICIENCY TEST REPORT  
Diag-11-2021-V0-EN**

**PROFICIENCY TEST FOR RABIES DIAGNOSIS**

**Session : 11-2021**

**Final report - Version 0**

**STUDY CODE : Diag-11-2021**

	<b>Name, First name</b>	<b>Function</b>	<b>Date</b>	<b>Signature</b>
<b>APPROVAL</b>	ROBARDET Emmanuelle	Coordinator	30/08/2021	
<b>AUTHORIZATION FOR DISTRIBUTION</b>	MONCHATRE-LEROY Elodie	LRFSN Director	30/08/2021	

This proficiency test report is distributed by Anses exclusively to proficiency test participants and the stake-holder. Anses declines the whole responsibility for the utilisation of this document by the holders, that are not allowed to use the provided data outside their internal use.

French Agency for Food, Environmental and Occupational Health Safety -  
Nancy Laboratory for Rabies and Wildlife -  
Technopole Agricole et Vétérinaire, Bâtiment H, CS 40 009, 54220 Malzéville - France  
Tél +33.3.83.29.89.50 – Fax : +33.3.83.29.89.58



## Table of contents

<b>1. INTRODUCTION</b>	<b>4</b>
<b>2. GENERAL INFORMATION</b>	<b>4</b>
2.1 IDENTIFICATION OF COORDINATOR AND STAFF INVOLVED IN THE STUDY	4
2.2 INSTRUCTION TO PARTICIPANTS	4
2.3 REGISTERED PARTICIPATING LABORATORIES	5
<b>3. PROFICIENCY TEST ITEMS</b>	<b>6</b>
3.1 PREPARATION OF THE PROFICIENCY TEST ITEMS	6
3.2 IDENTIFICATION OF THE PROFICIENCY TEST ITEMS	7
3.3 HOMOGENEITY	7
3.4 STABILITY	7
<b>4. LABORATORY PERFORMANCE EVALUATION</b>	<b>8</b>
<b>5. RESULTS</b>	<b>9</b>
<b>6. FOLLOW-UP OF INDIVIDUAL PERFORMANCES ON OVERALL DIAGNOSIS CONCLUSION</b>	<b>12</b>
<b>7. REFERENCES</b>	<b>13</b>
<b>8. ACKNOWLEDGMENTS</b>	<b>13</b>

## 1. INTRODUCTION

An inter-laboratory trial dedicated to rabies diagnosis was organized by the European Union Reference Laboratory (EURL) for Rabies. The objective was to assess the rabies diagnosis performance of laboratories based on recommended techniques as the Fluorescent Antibody Test (FAT) (OIE, 2018; WHO, 2018), the Rabies Tissue Culture Infection test (RTCIT) (OIE, 2018; WHO, 2018) and the biological molecular techniques represented by the conventional (OIE, 2018; WHO, 2018) and the Real Time RT-PCR (OIE, 2018; WHO, 2018). This work was undertaken in the frame of the Commission Regulation (EU) No 415/2013 of 6 May 2013 laying down additional responsibilities and tasks of the EURL for Rabies and amending Regulation (EC) No 737/2008 designating the EURL for Rabies.

## 2. GENERAL INFORMATION

### 2.1 IDENTIFICATION OF COORDINATOR AND STAFF INVOLVED IN THE STUDY

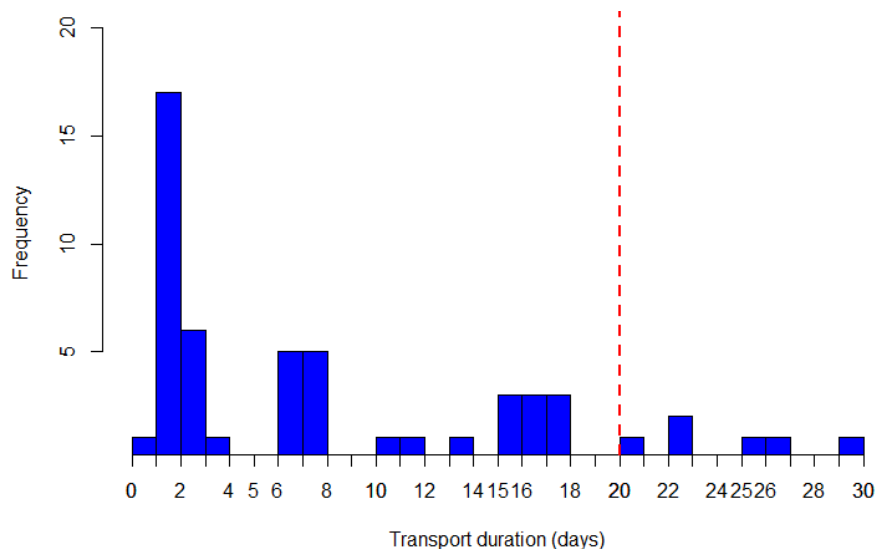
- **Director of the Laboratory:** E. Monchâtre-Leroy
- **EURL director and proficiency test Coordinator:** E. Robardet
- **Quality Manager:** F. Rizzo
- **Technical Staff:** J. Rieder and E. Longueval

### 2.2 INSTRUCTION TO PARTICIPANTS

The call for participation in the proficiency test for rabies diagnosis was sent to laboratories by e-mail in December 2020. At this step it was asked to participants to fill and sign the Anses Mutual Transfer Agreement, to frame work the exchange of strains and their utilisation and to take measure to have licence importation of samples ready for the transport, if requested, in accordance with the national custom clearance regulation.

A panel of 10 coded samples to be tested was sent to participating laboratories by a specialized carrier. Each sample contained 1ml of lyophilised homogenate of brains (animal origin) and was susceptible to be infected by a RABV, EBLV-1, EBLV-2, BBLV, DUVV or ABLV strain. The shipment of the panel to the laboratories started on May 31<sup>st</sup> 2021 at 4°C temperature by an international agreed carrier and under UN3373 conditions as specified by international regulation for the shipment of biological samples. All the panels were received by the participating laboratories during the time range where stability was ensured (20 days) except for six laboratories (L12; L26; L41; L44; L50; L56). Globally, 34% of the panels were received within 2 days, 47% within 6 days; 70% within 13 days and 89% within 20 days (Figure 1). Before its use, it was asked to laboratories to store the panel at 4°C as soon as received. Web links were sent to participants to fill online the acknowledgment (checking the satisfactory condition of reception of the samples) and the result forms.

**Figure 1:** Transport duration of panels for the 2021 session. Data based on 53 laboratories, according to their date reception statements, except for 4 laboratories for which carrier data was used.



Laboratories were requested to provide a unique diagnosis conclusion per sample according to their own rabies diagnosis procedure to mimic as much as possible routine diagnosis condition of participating laboratories. This diagnosis was the unique performance evaluation criteria of this proficiency test. The techniques considered to test the samples and for which the panel was validated were the following (s): FAT (Fluorescent Antibody test) and/or RTCIT (Rabies Tissue Culture Infection test) and/or conventional RT-PCR and/or Real Time RT-PCR. Expected diagnosis conclusion was a binomial response “positive” or “negative”.

Instruction to participants (FO-PQEILA-EXE-R\_INFORMATION-4) was sent in the same time than the call for participation sent in December. This document explained among others that the participant will not be evaluated in case of one or more missing value in the column positive or negative of the result form, in case of reception of the panel out of the limit of evaluated stability, or in case of result reception after the fixed deadline without prior to EURL agreement. The deadline for result reception was fixed on July 5<sup>th</sup> 2021.

### 2.3 REGISTERED PARTICIPATING LABORATORIES

In 2021, fifty-eight laboratories were registered in the inter-laboratory test for rabies diagnosis. Registered laboratories includes 25 National Reference Laboratories (NRLs) from the European Union (EU) and 33 laboratories from third countries (including 11 laboratories participating under EC fundings).

Forty-seven laboratories were registered for the FAT, 31 for the RTCIT, 32 for the conventional RT-PCR and 36 for the Real Time RT-PCR (Table 1). Proportion of registration for the different tests was 98% for the FAT, 53% for the RTCIT, 55% for the conventional RT-PCR and 62% for the Real Time RT-PCR.

**Table 1:** Number of laboratories registered as participants for the different techniques covered by 2021 rabies diagnosis inter-laboratory test.

	European Union NRLs	Third country laboratories participating under EC fundings	Third country laboratories participating not under EC fundings	Total
FAT	24	11	22	<b>57</b>
RTCIT	15	6	10	<b>31</b>
Conventional RT-PCR	16	4	12	<b>32</b>
Real Time RT- PCR	22	2	12	<b>36</b>
Total Laboratories	25	11	22	<b>58</b>

### 3. PROFICIENCY TEST ITEMS

#### 3.1 PREPARATION OF THE PROFICIENCY TEST ITEMS

The virus batches used in this study were produced by intra-cerebral inoculation of mice to reproduce as much as possible standard conditions of an animal infection. Viruses were produced according to the animal experimentation directives issued by the French Ethic Committee (Ethic committee approval number 17-076 and ministry agreement number #11772-2017101311312783). For each batch of virus, brains were excised after the death of the animals then mixed, homogenized, aliquoted into 1ml tubes and then freeze-dried. The positive samples of the panel were finally constituted of 7 blindly coded samples of freeze-dried homogenized brains infected with various rabies virus species.

The strains involved in the inter-laboratory test were:

- RABV Greece: a classical rabies virus (RABV) fox strain isolated in Greece in 2012.
- CVS 27: a RABV fixed strain.
- RABV dog Es: a RABV dog strain isolated in Spain in 2010.
- BBLV: a Bokeloh bat lyssavirus strain isolated in France in 2012.
- EBLV-1b: an European bat lyssavirus type 1 isolated in France in 2002.
- DUVV: a Duvenhage strain isolated in South Africa in 1971.
- Vaccinal Strain: a strain of vaccine induced case isolated in Latvia in 2014.

The material used as negative batch was negative brains of carnivores from French wild origin. An additional sample was also included in the panel test of each participant to avoid collusion. This sample differed from one laboratory to another and was originated from previous virus batches. These samples named "MIX" were not included in the evaluation process of the laboratories.

### 3.2 IDENTIFICATION OF THE PROFICIENCY TEST ITEMS

For each panel, all items were coded randomly. The code was constituted by the date of the inter-laboratory test campaign, the identification of the laboratory and the unique specific code of the item. Each item was dully labelled.

### 3.3 HOMOGENEITY

The evaluation of the homogeneity was undertaken for each positive batch by analysing in duplicate 10 randomly chosen samples.

- For all of them: FAT, RTCIT, Real Time RT-PCR SYBR Green was performed.
- For 3 samples, conventional RT-PCR and rapid genotyping by Taqman Real Time RT-PCR was also performed.
- For 1 sample, strain typing by sequencing was performed.

The evaluation of the homogeneity was undertaken for each negative batch by analysing in duplicate 10 randomly chosen samples by the following techniques: FAT, RTCIT, Real Time RT-PCR SYBR Green and conventional RT-PCR.

The analysis was performed after lyophilisation when all the samples were under their final form for the positive batches (RABV Greece, CVS27, RABV Dog Esp, BBLV, EBLV-1b, DUVV, Vaccinal Strain) and for the negative batches. All the batches were declared homogeneous as all the results were concordant to the expectations (positive samples were all found positives and negative samples were all found negatives whatever the technique used).

### 3.4 STABILITY

Stability of the panel was evaluated just before sending the panel under 4 different conditions:

- At day 0 (D0)
- After one week at Room Temperature (RT) (D6 and not D7 due to calendar constrains)
- After two weeks at RT (D14)
- After three weeks at RT (D21)

The samples were analysed with the FAT, the RTCIT and the Real Time RT-PCR SYBR Green to ensure that positive samples were positive and that negative samples were negative. The transport conditions requested to the specialised carrier being at +4°C, conditions tested were in consequence considered extreme situations. In the laboratory, lyophilised samples stored at +4°C have indeed been shown stable for 5 years.

Under the 4 conditions, all the positives samples were tested as positives and all negatives samples were tested as negatives.

Stability of the panel was consequently declared for 20 days of transport duration even with thawed ice-blocks conditions.

#### 4. LABORATORY PERFORMANCE EVALUATION

It was requested to laboratories to test the panel as usually done in their laboratory, by using their current own protocol to declare a rabies diagnosis conclusion for each sample. The rabies diagnosis conclusion per sample had to be expressed as a binomial response “positive” or “negative”.

For the rabies diagnosis performance evaluation, correct results on the overall diagnosis conclusion were required for each sample meaning that the evaluation was considered as “satisfactory” when the participating laboratory declared the rabies diagnosis conclusion corresponding to the determined status of the samples (positive or negative) while the evaluation was considered “unsatisfactory” when at least discordant result occurs.

A performance score was attributed to the laboratories according to the number of discordant results as followed:

- No discrepant results: Score 1 = Satisfactory performance
- 1 discrepant result : Score -1 = Unsatisfactory performance
- 2 discrepant results: Score -2 = Unsatisfactory performance
- 3 discrepant results: Score -3 = Unsatisfactory performance
- 4 discrepant results: Score -4 = Unsatisfactory performance
- 5 discrepant results: Score -5 = Unsatisfactory performance
- 6 discrepant results: Score -6 = Unsatisfactory performance
- 7 discrepant results: Score -7 = Unsatisfactory performance
- 8 discrepant results: Score -8 = Unsatisfactory performance
- 9 discrepant results: Score -9 = Unsatisfactory performance



## 5. RESULTS

Fifty laboratories provided answer to the overall rabies diagnosis assessment (Table 2 and table 4).

Six laboratories experienced long transport duration out of the range where stability was declared (>20 days). As stated in the proficiency test instruction FO-PQEILA-EXE-R\_INFORMATION-4, the performance of those laboratories was not evaluated (**L12; L26; L41; L44; L50; L56**).

On the 44 evaluated laboratories, two laboratories (**L09** and **L31**) (5% of evaluated laboratories) provided at least one discrepant result: a false positive diagnosis conclusion on a negative sample (**L09** and **L31**) and a false negative conclusion on a CVS 27 positive sample (**L31**).

Table 2: Overall diagnosis conclusion results per strain

	n laboratories with discrepant results/ total laboratories	% of laboratories with discrepant results and 95 CI
Total	2/44	4.5 [0.6 – 15.5]
Negative samples	2/44	4.5 [0.6 – 15.5]
Positive samples	1/44	2.3 [0.1 – 12.0]
RABV samples	1/44	2.3 [0.1 – 12.0]
CVS 27	1/44	2.3 [0.1 – 12.0]
RABV Greece	0/44	0 [0.0 – 8.0]
RABV Dog Es	0/44	0 [0.0 – 8.0]
Vaccinal strain	0/44	0 [0.0 – 8.0]
EBLV-1	0/43	0 [0.0 – 8.2]
BBLV	0/42	0 [0.0 – 8.4]
DUVV	0/43	0 [0.0 – 8.2]

Individual laboratory scores for rabies diagnosis and corresponding performance evaluations are indicated in the table 3.

Table 3: Rabies diagnosis performance evaluation per laboratory. NA: Not applicable.

Lab code	Score	Evaluation
L02	1	Satisfactory
L03	1	Satisfactory
L05	1	Satisfactory
L06	1	Satisfactory
L07	1	Satisfactory
L08	1	Satisfactory
<b>L09</b>	<b>-1</b>	<b>Unsatisfactory</b>
L10	1	Satisfactory
L11	1	Satisfactory
L12	NA	Not evaluated
L13	1	Satisfactory
L14	1	Satisfactory
L15	1	Satisfactory
L16	1	Satisfactory
L17	1	Satisfactory
L18	1	Satisfactory
L20	1	Satisfactory
L21	1	Satisfactory
L22	1	Satisfactory
L23	1	Satisfactory
L24	1	Satisfactory
L25	1	Satisfactory
L26	NA	Not evaluated
L27	1	Satisfactory
L28	1	Satisfactory
L29	1	Satisfactory
L30	1	Satisfactory
<b>L31</b>	<b>-2</b>	<b>Unsatisfactory</b>
L32	1	Satisfactory
L33	1	Satisfactory
L34	1	Satisfactory
L35	1	Satisfactory
L36	1	Satisfactory
L37	1	Satisfactory
L38	1	Satisfactory
L40	1	Satisfactory

Lab code	Score	Evaluation
L41	NA	Not evaluated
L42	1	Satisfactory
L44	NA	Not evaluated
L46	1	Satisfactory
L48	1	Satisfactory
L49	1	Satisfactory
L50	NA	Not evaluated
L51	1	Satisfactory
L52	1	Satisfactory
L54	1	Satisfactory
L55	1	Satisfactory
L56	NA	Not evaluated
L57	1	Satisfactory
L58	1	Satisfactory



## 6. FOLLOW-UP OF INDIVIDUAL PERFORMANCES ON OVERALL DIAGNOSIS CONCLUSION

Within the last three sessions, where overall diagnosis conclusion evaluation was assessed, the number of laboratories presenting an unsatisfactory evaluation vary from 1 to 2 per year (1 laboratory in 2017; 2 in 2019 and 2 in 2021). Unsatisfactory evaluation was an isolated event as none of the evaluated laboratories repeated this event. Laboratories with unsatisfactory evaluation in 2021 are laboratories that did not participate the last two years (Table 5).

Table 5: Performance evaluation evolution of participating laboratories in the 2021 session. S: Satisfactory evaluation; UNS: Unsatisfactory evaluation.

Laboratory code 2021	2017	2019	2021
L02		S	S
L03	S	S	S
L05	S	S	S
L06	S	S	S
L07			S
L08	S	S	S
L09			UNS
L10		S	S
L11	S	S	S
L13	S	S	S
L14	S	S	S
L15	S	S	S
L16	S	S	S
L17	S	S	S
L18	S	S	S
L20	S	S	S
L21	S	S	S
L22	S	S	S
L23	S	S	S
L24	S		S
L25			S
L27	S	S	S
L28	S	S	S
L29	S	S	S
L30	S	S	S
L31			UNS
L32	S	S	S
L33	S	S	S
L34			S
L35	S	S	S

Laboratory code 2021	2017	2019	2021
L36	S	S	S
L37	S	S	S
L38	S		S
L40	S	S	S
L42	S		S
L46	S		S
L48	S	S	S
L49	S	S	S
L51	S	S	S
L52	S	S	S
L54	S	S	S
L55	S	S	S
L57	S	S	S
L58	S	S	S

## 7. REFERENCES

OIE, 2018, *Manual of standards for diagnostic tests and vaccines, Rabies, Chapter 3.1.17.* 35 p.

[http://www.oie.int/fileadmin/Home/eng/Health\\_standards/tahm/3.01.17\\_RABIES.pdf](http://www.oie.int/fileadmin/Home/eng/Health_standards/tahm/3.01.17_RABIES.pdf)

World Health Organization, Rupprecht, Charles E, Fooks, Anthony R & Abela-Ridder, Bernadette. (2018). *Laboratory techniques in rabies, volume 1, 5th ed.* World Health Organization.

<https://apps.who.int/iris/handle/10665/310836>. Licence: CC BY-NC-SA 3.0 IGO. 289 p.

## 8. ACKNOWLEDGMENTS

This study was funded by the European Commission and by ANSES.

We would like to thank the ANSES staff involved in this study for carrying out the technical work:

- panel constitution and result reception: J. Rieder and E. Longueval;
- experiment on mice and rabies diagnosis: V. Brogat, S. Kempff and E. Litaize under the management of A. Servat;
- molecular biology: M. Badre-Biarnais, C. Peytavin and J.L. Schereffer under the management of E. Picard-Meyer.

We would like also to thank scientists who kindly provided the strains used for the trial.